**Title**

An experimental framework for determining the degree of intraguild predation in a three-species terrestrial omnivorous arthropod food web in the field

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**Abstract**

1. Intraguild predation (IGP) is common in natural and human-managed systems and plays a critical role in food web dynamics. Previous studies have documented the occurrence of IGP across a wide range of arthropod predator taxa, yet there is still a lack of quantitative understanding regarding the degree/intensity of IGP in these systems.
2. I propose an experimental framework combining controlled feeding trials and stable isotope analysis to determine the degree of IGP in a three-species terrestrial omnivorous arthropod food web (shared prey + mesopredator + top predator) in the field. The degree of IGP is defined herein as the proportion (in number) of mesopredator consumed in the total diet (shared prey + mesopredator) of top predator. The feeding trials are used to construct a standard curve for the degree of IGP in the focal system, to which the stable isotope signatures of field samples are compared to estimate the degree of IGP in the field.
3. The proposed framework leverages the strengths of different experimental approaches to studying trophic interactions, providing a useful tool for quantifying IGP in a more accurate (controlled feeding trials and standard IGP curve) and realistic (stable isotope analysis of field samples) fashion.
4. If proven successful, the current framework can be extended to food webs involving more complex interactions (e.g., cannibalism and multiple shared prey) and further complemented with other approaches (e.g., molecular gut content analysis) to capture a more complete picture of IGP dynamics in the field.

**Key words**

intraguild predation, food webs, terrestrial arthropods, stable isotope analysis, trophic interactions, feeding experiment

**Introduction**

Intraguild predation (IGP) is common in natural and human-managed ecosystems (Polis & Holt, 1992; Müller & Brodeur, 2002; Arim & Marquet, 2004) and has been documented across a wide range of arthropod predator taxa (Polis et al., 1989; Gagnon et al., 2011). IGP could substantially affect the abundance and distribution of interacting species, generating ecological and evolutionary consequences for food web dynamics (Polis et al., 1989).

Previous studies have revealed the occurrence of IGP among arthropod predators through field observations of their diet compositions (e.g., Nyffeler & Sunderland, 2003; Birkhofer & Wolters, 2012). Manipulative experiments (e.g., field cages) have also been used to assess the intensity of IGP (Denno et al., 2004; Provost et al., 2005), which could reveal the mechanisms underlying predator-prey interactions and allow for strong causal inferences about IGP. However, the confined settings in these experiments may potentially alter the encounter rates between organisms, thus leading to biased results (Uiterwaal et al., 2019).

Stable isotopes, particularly nitrogen isotope signatures (δ15N), have been used to estimate the trophic level of predators in the field for inferences about IGP (Halaj et al., 2005; Wise et al., 2006; Sanders & Platner, 2007). It is suggested that IGP would increase the δ15N of predators (Ponsard & Arditi, 2000). Rickers et al. (2006) conducted feeding experiments on wolf spiders (*Alopecosa cuneata*) and revealed a higher δ15N of these top predators in IGP treatment. However, the study did not quantify the degree of IGP as the IGP treatment was binary (absence vs. presence of mesopredator) with constant numbers of shared prey and mesopredator. On top of that, the trophic levels of top predators in previous studies were often calculated based on assumed trophic discrimination factors (e.g., Klarner et al., 2013; Svanbäck et al., 2015). Since trophic discrimination factors are quite taxon-specific (Caut et al., 2009), this could lead to incorrect trophic level estimates and thus inferences about IGP in the field.

Recently, researchers have applied molecular gut content analysis and immunological techniques to reliably detect the presence of certain food items in predators’ diet (Hagler, 2006; Gagnon et al., 2011; Mansfield & Hagler, 2016), allowing for estimating the incidence rate of IGP (i.e., the percentage of top predator individuals with mesopredator detected in the gut contents). Nonetheless, the incidence rate may not necessarily reflect the degree of IGP in the system (Raso et al., 2014). For example, it is possible that a high percentage of top predator individuals feed on mesopredator yet each top predator individual consumes on average a low proportion of mesopredator in the diet. In this case, a high incidence rate of IGP would be misleading and fail to capture the overall IGP dynamics (including frequency and intensity).

So far, there is still a lack of quantitative information regarding the intensity/degree of IGP in the field, and such information is the first step towards a deeper understanding of food web dynamics [allometric constraints?]. To address this gap, I propose an experimental framework combining controlled feeding trials and stable isotope analysis of field samples to determine the degree of IGP in a three-species terrestrial omnivorous arthropod food web (top predator + mesopredator + shared prey). The degree of IGP is defined herein as the proportion (in number) of mesopredator consumed in the total diet (mesopredator + shared prey) of top predator. The feeding trials will experimentally link different levels of mesopredator consumption by top predators to the changes in their nitrogen isotope signatures (δ15N) via a standard curve, to which the isotope signatures of field samples are compared to estimate the degree of IGP in the field.

**Materials and Methods**

Here, I illustrate the proposed framework using an example of a terrestrial arthropod food web involving a spider top predator, a spider mesopredator, and a planthopper shared prey (Fig. 1a). Two sets of controlled feeding trials will be conducted. In the first trial, the top predator and the mesopredator are fed the shared prey (Fig. 1b). As invertebrates may exhibit rapid tissue turnover, a period of 5–10 days will allow predators to incorporate isotope signatures into their tissues and reach an isotopic equilibrium state with the shared prey (Quinby et al., 2020). In the second trial (same duration as the first trial), the top predator is fed mixed diets with different proportions of shared prey and mesopredator individuals from the first trial: (1) shared prey only, (2) 75% of shared prey + 25% of mesopredator, (3) 50% of shared prey + 50% of mesopredator, (4) 25% of shared prey + 75% of mesopredator, and (5) mesopredator only (Fig. 1c). The numbers of shared prey and mesopredator in the supplied diet can be determined based on their field densities. The purpose of the second trial is to simulate a full range of potential encounter rates that the focal organisms might experience in the field.

At the end of the second trial, the numbers of shared prey and mesopredator consumed by the top predator in each diet treatment are recorded, and the difference in nitrogen isotope signatures between the top predator individuals and the shared prey (baseline) are analyzed (δ15N*predator*─ δ15N*prey*; Δ15N). A standard curve can be constructed by plotting the experimental Δ15N of top predator against the proportion of mesopredator consumed (Fig. 1d). Finally, field samples of shared prey and top predator individuals are collected, with their δ15N analyzed to obtain the empirical Δ15N. The degree of IGP in the field can thus be estimated by interpolating the empirical Δ15N to the standard curve (Fig. 1e). A hypothetical example of data collection in the second trial for standard curve construction is provided in Fig. 2.

**Results and Discussion**

The proposed experimental framework leverages the strengths of previous approaches to studying IGP—the controlled feeding trials combined with stable isotope analysis can yield accurate experimental Δ15N to construct a standard curve, whereas the empirical Δ15N derived from stable isotope analysis of field samples reflects the trophic interactions under natural settings. Together, this framework provides a useful tool for determining the degree of IGP in the field in a more quantitative and realistic fashion.

Agricultural systems, in which IGP has been frequently documented (Polis et al., 1989; Rosenheim et al., 1995), are ideal for testing the proposed framework. The food web structures in such systems are relatively simple compared with other ecosystems. Therefore, the potential confounding effects of non-focal species on the trophic interactions among focal organisms can be minimized (Vance-Chalcraft et al., 2007). Moreover, this framework is useful for quantifying the effects of various abiotic and biotic factors on IGP under field settings. For instance, the framework can be implemented along a gradient of habitat complexity to examine how different levels of habitat complexity might affect the degree of IGP in the field. Finally, the framework is robust to variations in background isotope signatures because the degree of IGP is determined based on the difference between nitrogen isotope signatures of the focal organisms (Δ15N) rather than their original values (δ15N), thus allowing for comparisons across sites or systems with distinct background isotope signatures.

A better quantitative understanding of IGP can provide critical insights into the complex predator-predator-prey trophic interactions and could help predict the community structure and stability (Arim & Marquet, 2004; Nakazawa & Yamamura, 2006; Pahl et al., 2020). Furthermore, such an understanding can have useful implications for practitioners, for example, evaluation of the effectiveness of biocontrol agents in pest control programs (Müller & Brodeur, 2002). If proven successful, the current framework can be further extended to food webs involving more complex interactions (e.g., cannibalism and multiple shared prey) and complemented with other approaches (e.g., molecular gut content analysis) to better elucidate the IGP dynamics in the field.

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**Contribution of authors**

GCH conceived the idea and wrote the manuscript. No other person was entitled to authorship.

**[20 references at max]**

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**Things to revise:**

* Check the suggested citations and trim the unnecessary ones down to 20
* Check the “allometric constraints”
* Narrow the scope and focus on terrestrial arthropod predators
* Address the concerns of generalist predators’ diet in the field: N values not just simply reflecting the mixing of two prey but also secondary predation, scavenging or the fact that predators consume a very diverse mix or prey over their life time

**Figure legends**

Figure 1.A schematic diagram of the proposed experimental framework for determining the degree of intraguild predation (IGP) in a three-species terrestrial omnivorous arthropod food web involving a spider top predator, a spider mesopredator, and a planthopper shared prey (a). In the first trial (b), the top predator and the mesopredator are fed the shared prey for an appropriate time period to ensure an isotopic equilibrium state between predators and the shared prey. In the second trial (c), the top predator is fed mixed diets with different proportions of shared prey and mesopredator individuals to simulate a full range of potential encounter rates that the focal organisms might experience in the field. (d) A standard curve can be constructed by plotting the difference in nitrogen isotope signatures between the top predator individuals and the shared prey (baseline) (δ15N*predator*─ δ15N*prey*; experimental Δ15N) against the proportion of mesopredator consumed. Note that the curve may not necessarily be linear due to the differences in the biomass of shared prey and mesopredator individuals. (e) The δ15N of field-sampled shared prey and top predator individuals are analyzed to obtain the empirical Δ15N, which is then interpolated to the standard curve to estimate the degree of IGP in the field.

Figure 2. A hypothetical example of data collection in the second trial for standard curve construction. Each diet treatment consists of five replicates (i.e., five different top predator individuals). *N*: number of shared prey/mesopredator supplied in the mixed diet; *C*: number of shared prey/mesopredator consumed by the top predators; *P*: proportion of mesopredator consumed (%). Each point in the standard curve represents a top predator individual.

**Figures**

Figure 1.

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Figure 2.

